
1 Micro-Evo-Devo

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13 Abstract

14 Micro-evo-devo **as** the study of genetic basis of developmentally mediated
15 phenotypic variation, and the evolutionary forces that affect the fate of that
16 variation within populations. Currently, there are few studies detailed enough to
17 trace the short-term evolutionary events underlying changes in developmental
18 processes, which leaves the general explanatory power of a micro-evo-devo
19 approach unclear. One promising approach is to use artificial selection to directly
20 cause evolution of developmentally mediated phenotypes and then infer the
21 genetic and developmental underpinnings of that response. A second promising
22 approach is to use a comparative approach in model systems such as sticklebacks
23 that have repeatedly been challenged to adapt to similar environments in the
24 recent past. Part of the reason that micro-evo-devo studies are still rare is an
25 implicit assumption by some that the current variability of a population is
26 irrelevant to long-term patterns, and that the vast majority of short-term changes
27 in development are essentially random with respect to long-term trends. On the
28 other hand, all major evolutionary transitions or trends must consist of micro-

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29 evolutionary changes. Emerging evidence from a variety of systems shows that
30 long-term patterns can sometimes be retrodicted from variability within extant
31 populations. Further development of micro-evo-devo approaches is needed to
32 enable us to determine the generality of these results.

33 Keywords

34 Genotype-phenotype-fitness map · Micro-evo-devo · Evolve and resequence ·
35 Variability · Quantitative genetics

36 Introduction

37 Understanding the evolution of phenotypic diversity is an enduring goal of biology.
38 Two complementary approaches are necessary to realize that goal: comparative and
39 process-based. The comparative approach describes biological diversity among taxa
40 from the genomic to the phenotypic level, and places that diversity in a phylogenetic
41 context. The process-based approach investigates how traits that exemplify that
42 diversity actually evolve within populations (see the chapter ▶ [“A Process-Based
43 Approach to the Study of Flower Morphological Variation”](#)).

44 The increasing importance of evolutionary developmental biology, or evo-devo,
45 comes from the fact that much of that biological diversity is generated by develop-
46 mental processes. The field of evo-devo arose from two nearly coincident develop-
47 ments in biology in the late 1970s and 1980s (Laubichler 2010). At this time, a
48 number of evolutionary biologists reemphasized the explanation of the evolution of
49 phenotypes rather than genes as their goal (see the chapter ▶ [“The Role of Evo-Devo
50 in the Extended Synthesis”](#) for a different perspective on this shift). Nearly simul-
51 taneously, molecular developmental geneticists scored their first dramatic successes
52 in identifying genes responsible for fundamental differences in development among
53 taxa. Since then the core work in evo-devo has remained comparative, with a focus
54 on identifying both the fundamental processes that underpin development in multi-
55 ple taxa, and the underlying developmental and genetic basis of discrete differences
56 among species. These approaches only indirectly inform us about the processes that
57 generated developmental diversity.

58 This chapter briefly reviews the current status of the process-based approach in
59 the evo-devo studies. Despite early calls for the process-based study of evo-devo
60 (Stern 2000; Johnson and Porter 2001), such studies are still relatively uncommon.
61 This leaves us uncertain whether the micro-level processes by which development
62 must evolve influence the larger-scale events and patterns that can be studied using
63 the comparative approach.

64 What Is Micro-Evo-Devo?

65 We define micro-evo-devo as the study of the nature and genetic basis of develop-
66 mentally mediated phenotypic variation, and the evolutionary forces that affect the
67 fate of that variation within populations. These twin emphases on the nature of
68 phenotypic variation and on the interaction between that variation and natural
69 selection, the genotype-phenotype-fitness map, are what make micro-evo-devo
70 studies distinct from their parent disciplines, population, and quantitative genetics.
71 The genotype-phenotype (GP) map (see the chapter ▶ [“Genotype-Phenotype Map”](#))
72 is the biological machinery that turns a genotype into all the many phenotypes that an
73 individual expresses during its lifetime (Lewontin 1974). By extending the map from
74 phenotypes to fitness, one can in principle include all of the evolutionary forces that
75 are hypothesized to give rise to properties of the GP map that generate the evolu-
76 tionary properties of developmental systems, including evolvability (see the chapters
77 ▶ [“Evolvability”](#) and ▶ [“Variational Approaches to Evolvability: Short- and Long-
78 Term Perspectives”](#)), modularity (see the chapter ▶ [“Modularity in Evo-Devo”](#)),
79 plasticity (see the chapters ▶ [“Developmental Plasticity and Evolution”](#) and
80 ▶ [“Eco-Evo-Devo”](#)), robustness, etc. Today micro-evo-devo encompasses studies
81 of mutation and standing variation that affect development, the responses of mor-
82 phology, and the developmental processes that underlie it to natural and artificial
83 selection, and the causes of natural selection on morphological traits. These
84 approaches are related to and complement both the “devo-evo” approach to the
85 study of how developmental processes shape the evolutionary process (see chapter
86 ▶ [“Developmental Evolutionary Biology \(Devo-Evo\)”](#)) and comparative studies of
87 recently diverged populations or species where contemporary data on variation and
88 selection is relevant to the divergence under study.

89 Traditional evo-devo can address the nature of the genetic and developmental
90 differences that have given rise to biological diversity, but cannot directly answer
91 questions about why that particular variation has been recruited to evolutionary
92 divergence. For example, a finding that a particular difference in morphology is
93 the result of changes in gene regulation tells us what must have evolved – the
94 cis-regulatory elements of genes involved in development – but it does not tell us
95 how or why they evolved that way. Are there alternative paths that evolution might
96 have taken? If so, why weren’t they? Only at the population level can we hope to
97 study all the evolutionarily relevant variation and its phenotypic effects.

98 While evolutionary biologists have long realized that the study of the evolution of
99 development must ultimately encompass the origin and fixation of genetic variants
100 affecting development, it took longer for the new molecular tools of developmental
101 genetics to become sufficiently inexpensive for population-level studies. Explicit
102 micro-scale studies of evo-devo are still relatively uncommon (Nunes et al. 2013).
103 While some of this relative neglect is based on the historical origins of evo-devo
104 outlined above, we suspect that it is also due to an implicit assumption among at least
105 some evo-devo practitioners that what happens at the micro scale is not very relevant
106 to the differences among species and higher taxa (e.g., Milocco and Salazar-Ciudad
107 2020). The use of the term “micro” implies an overlap with the more general micro-

108 vs. macro-evolution distinction which concerns the relationship between evolution
109 over short vs. long time scales or that within vs. among species. The skepticism that
110 micro-evo-devo is relevant inherits some of aspects of the controversy over whether
111 macro-evolution is just micro-evolution continued over a long time period (e.g.,
112 Gould 2002). Currently the connections between micro-evolution of development
113 and the macro-pattern of diversity in development are unclear, as there are limited
114 number of relevant studies. The key challenge is to either establish the relevance of
115 micro-evo-devo to the larger issues in evolutionary developmental biology, or,
116 alternatively, explain how larger-scale or longer-term processes render micro-level
117 changes irrelevant.

118 **Micro-Evo-Devo in Action**

119 To illustrate the scope of micro-evo-devo studies, let's consider example studies that
120 span the diversity of approaches that are possible.

121 The most direct micro approach is to observe the response of a population to
122 selection on a developmentally relevant phenotype. Selection can be used in several
123 contexts. Natural populations can frequently experience selection, but it is rarely
124 clear what the actual target of selection might be. In artificial selection the investi-
125 gator measures and selects the individuals chosen for breeding. Artificial selection
126 experiments are thus ideal for testing hypotheses about the nature of the develop-
127 mental response when a particular phenotypic change is selected for.

128 For example, Marchini and Rolian (2018) investigated the mechanism by which a
129 mouse population responded to selection for tibia length, one of the long-bones that
130 has diversified among mammals. After only 14 generations of artificial selection for
131 increased tibia length standardized by body mass, they had increased average length
132 by 9% and 14% in 2 replicate populations. During bone development, a scaffolding
133 of cartilage is produced by chondrocytes at each end of the developing bone,
134 undergoes proliferation and enlargement, and then that scaffolding is filled in by
135 the osteoblasts that form ossified, mineralized bone. Based on this knowledge,
136 Marchini and Rolian (2018) could identify four potential mechanisms for an evolved
137 increase in tibia length: prolongation of the period of bone growth, increases in the
138 number of dividing chondrocytes, increases in chondrocyte size, and increases in the
139 proliferation rate during ossification. Analysis of mice at the end of the experiment
140 showed that the long-limbed treatments had increased the number of chondrocytes,
141 while no differences were found in the other three characteristics. In this population,
142 evolution did not proceed by all possible means, but was focused on just one process.
143 This is in contrast with the differences in growth between limbs in bats, which have
144 elongated forelimbs, and jerboas, which have elongated hindlimbs. In these taxa,
145 increases in chondrocyte size are important in the elongated body parts. This
146 difference in mechanism may reflect the fact that the experimental mice were
147 selected to increase the length of one limb, but not to change the relative growth
148 of their limbs.

149 Other micro-evo-devo studies investigate the genetic changes underlying changes
150 in development, rather than directly studying development. In an evolve-and-
151 resequence (E&R) experiment, next-generation sequencing of selected populations
152 identifies the genomic changes that distinguish the populations after selection.
153 Changes that cannot be explained by drift are inferred to be due to selection. For
154 example, Turner et al. (2011) selected replicate populations of *Drosophila*
155 *melanogaster* to have larger or smaller body size for 100 generations. The genomic
156 response was traceable to a large number of genomic regions. The regions that
157 responded to selection were far more likely to code for genes involved in the gene
158 ontology categories anatomical development, cell number, and metamorphosis. This
159 suggests that the changes in size in this experiment were accomplished by small
160 alterations in a large number of developmental processes.

161 Studies of mice and flies like those just mentioned take advantage of the fact that
162 these are premier model systems for the study of development, but are hampered by
163 the difficulty of studying natural populations. In contrast, *Gasterosteus aculeatus*,
164 the three-spine stickleback was chosen for micro-evo-devo studies due to a wealth of
165 recently differentiated natural populations. This fish and its close relatives are found
166 in near-shore marine environments in the Northern Hemisphere. They have repeat-
167 edly invaded freshwater habitats from the oceans, particularly as deglaciation pro-
168 ceeded after the last ice age, making every river and stream an independent
169 evolutionary experiment with a geologic time signature. Sticklebacks have conse-
170 quently been a favorite subject for studies of phenotypic evolution (Bell and Foster
171 1994), including the genetic basis of developmental changes. For micro-evo-devo,
172 these natural experiments provide many of the advantages of artificial selection
173 experiments, plus the fact that nature has done the work of selection over a much
174 longer time period than is practical in the laboratory. For many traits the resulting
175 differences in phenotypes between founder and derived populations are large enough
176 to make it possible to use the comparative evo-devo toolkit to identify and test
177 hypotheses about the basis of adaptation. This combination of features enables
178 investigators to compare variation in the marine founder population to the products
179 of evolution based on that variation, while investigating the cause of those
180 differences.

181 For example, many freshwater populations undergo loss of spines and armor that
182 help protect marine fish from predation. In freshwater these features actually increase
183 predation by insect predators, and increase the demand for calcium, which is far
184 more limited in freshwater habitats. In addition to contemporary studies
185 documenting these selective forces, detailed fossil sequences show that the loss of
186 spines and armor takes place gradually over a few 1000 generations (Hunt et al.
187 2008). Genetic changes that make large contributions to these repeated losses have
188 been traced to cis-regulatory changes at two key developmental genes. The armor
189 plates are greatly reduced by a mutation in the regulatory region of the *ectodysplasin*
190 (*EDA*) gene (Colosimo et al. 2005; O’Brown et al. 2015), which plays an important
191 role in cell-signaling during developmental of the neural crest and ectoderm.
192 Remarkably, the same allele (haplotype) is involved in this adaptation throughout
193 much of the range of the species. It is maintained at low frequency in the marine

194 population which founds each freshwater population by gene flow from the existing
195 freshwater populations. This enables rapid adaptation upon invasion of a new
196 drainage. In contrast, the genetic basis for loss of pelvic spines in some freshwater
197 populations has been traced to deletions of an enhancer at the *Pituitary homeobox 1*
198 (*Pitx1*) locus (Chan et al. 2010). In this case, the deletion present in each drainage
199 was unique. Several lines of evidence suggest that this enhancer is in a region of the
200 genome with a high mutation rate for deletions. Thus, in both of these cases of armor
201 loss, the supply of the variation is enhanced, potentially explaining why these two
202 regions are responsible for parallel adaptive events.

203 These dramatic cases of evolution due to fixation of large effect alleles do not
204 explain all aspects of stickleback adaptation. Studies that map the genetic differences
205 underlying many differentiated traits show that such changes typically involve a
206 large number of genetic changes with a range of effects sizes (Peichel and Marques
207 2017). Even in the case of armor loss, many other genomic regions contribute to the
208 multifarious differences among populations. For example *Eda* has a large effect on
209 plate number, but minor effects on the size of the plates that are produced. A very
210 useful heuristic model for adaptation is that of Fisher's geometric model (Orr 2005)
211 of simultaneous adaptation of many traits. Fisher assumed that each trait has an
212 optimal state at which fitness is maximized, and that the fitness of a phenotype is a
213 smooth function of the distance to that optimum in the space of all possible
214 phenotypes. He also assumed "universal pleiotropy," meaning that each mutation
215 has some effect on all traits, although the combination of effects differs among
216 mutations. When the population is not at the optimum, mutations will increase
217 fitness when they move the phenotypic state closer to the optimum, and otherwise
218 will decrease fitness. When the population is far from the optimum, mutations with
219 large effects can be favored, but as the population approaches the optimum, variants
220 with large effects are no longer favored, even if they would have provided a better
221 initial step towards the optimum. Instead, a series of smaller effect mutations are
222 fixed as the population approaches the optimum. The pattern of effects sizes in the
223 adaptation of stickleback traits matches the expectation under Fisher's geometric
224 model well (Peichel and Marques 2017).

225 **Building Blocks of Micro-Evo-Devo**

226 Many comparisons of evo-devo with traditional micro-evolutionary biology con-
227 sider the latter to be synonymous with population genetics. The purview of popula-
228 tion genetics is generation and inheritance of discrete inherited variants in their
229 genetic context and the study of the forces affecting the fates of those genetic
230 variants within populations. The phenotype plays no necessary role in population
231 genetics once the relative fitness of genotypes is specified, although many specific
232 population genetics models do include a connection between fitness and phenotype,
233 albeit in a usually simplified and rather abstract form. This phenotype-blindness of
234 population genetics has been the subject of scorn not only from practitioners of

235 evo-devo but also from biologists more concerned with the evolution of phenotypes
236 than genes (Laubichler 2010).

237 There is, however, another tradition of micro-evolutionary studies based on
238 quantitative traits that is complementary to that of population genetics. The field of
239 quantitative genetics infers the inheritance of phenotypes from phenotypic data on
240 related individuals without directly assuming anything about the underlying genetic
241 variants. The nature of the phenotypic similarities allows the quantitative geneticist
242 to predict the response of phenotypes to natural and artificial selection. Quantitative
243 genetics is a natural way to evaluate important aspects of genetic architecture, such
244 as evolvability (see the chapters ▶ “Evolvability” and ▶ “Variational Approaches to
245 Evolvability: Short- and Long-Term Perspectives”), modularity (see the chapter
246 ▶ “Modularity in Evo-Devo”), and canalization (see the chapter ▶ “Canalization:
247 A Central But Controversial Concept in Evo-Devo”). The generation of new phe-
248 notypic variation by mutation or as a plastic response to the environment is also in
249 the purview of quantitative genetics (e.g., Braendle et al. 2010; Houle and Fierst
250 2013; see also the chapter ▶ “Developmental Plasticity and Evolution”).

251 An important aspect of this quantitative arm of micro-evolutionary biology is that
252 the study of inheritance and selection readily generalizes to the study of complex
253 phenotypes consisting of many potentially interrelated parts (Lande 1979; Lande and
254 Arnold 1983). Almost any morphological phenotype is complex in this sense.
255 Unfortunately, many studies of the evolution of development at all scales make the
256 unrealistic but convenient assumption that phenotypes can be represented by a single
257 measurement or as a set of discrete alternative states. The ability to measure complex
258 phenotypes is increasing rapidly (see the chapter ▶ “Phenotyping in Evo-Devo”),
259 and incorporating these multivariate data is a challenge for all aspects of evo-devo
260 studies (e.g., Pitchers et al. 2019).

261 Despite capturing largely complementary aspects of evolution, the population
262 genetic and quantitative genetic approaches unfortunately cannot readily be com-
263 bined into a single comprehensive approach to micro-evolution. They use
264 completely dissimilar formalisms where the parameters in one tradition have no
265 equivalents in the other tradition. For example, population genetics assign fitnesses
266 to discrete genotypes, but discrete genotypes are not identified in the quantitative
267 tradition. The fitness functions estimated in the quantitative tradition cannot be
268 applied to genetic variants that lack phenotype information.

269 Building GP Maps

270 Population genetics and quantitative genetics operate on opposite sides of the GP
271 map (Lewontin 1974). It has long been clear that what we need to fuse the
272 complementary advantages of population and quantitative genetics is a GP map.
273 Micro-evo-devo studies necessarily incorporate both population genetic and quan-
274 titative genetic concepts, connected through the concept of the GP map (see the
275 chapter ▶ “Genotype-Phenotype Map”). Key advantages of a GP map-based
276 approach include gaining a handle on the empirically challenging concept of

277 pleiotropy (see the chapter ► [“Pleiotropy and Its Evolution: Connecting Evo-Devo](#)
278 [and Population Genetics”](#)). Without an a priori notion of the map, the only way to
279 study pleiotropy is to estimate effects of genetic variants on the prohibitively large
280 universe of possible traits affected (Pitchers et al. 2019). Furthermore, the GP map
281 concept enables study of how the map itself can evolve (see the chapter on ► [“Epis-](#)
282 [tasis”](#)), potentially reshaping evolutionarily important parameters, including pleiotro-
283 pty and evolvability.

284 The difficulty is that, with a few limited exceptions, GP maps are very poorly
285 known. What knowledge evolutionary biologists have of GP maps usually comes
286 from elsewhere in biology – from mutational screens, and from cell and develop-
287 mental biology. Often such information initially is no more than a pointer for future
288 work “Look in or around this gene for relevant genetic variation.” In addition, there
289 are many vaguer generalizations that we use as proxies for parts of the GP map, such
290 as “premature stop codons have large phenotypic effects” and “synonymous amino
291 acid substitutions will usually have no phenotypic effects.”

292 Traditional developmental and evo-devo studies often build out parts of
293 genotype-phenotype map as a byproduct of researching the function or evolution
294 of genes or pathways. For example, the transcription factor *ovo* in *Drosophila*
295 *melanogaster* and closely related species (reviewed in Chapter 6 of Wagner 2014)
296 has 3 alternative promoters, 5 different transcripts with different functions and at
297 least 13 enhancers. One transcript is necessary in primordial germ cells, while that
298 and a second transcript act antagonistically to regulate gene expression in the ovary.
299 A third transcript is involved in several aspects of cuticle differentiation, and three
300 enhancers of this transcript generate spatially precise effects on the placement of
301 trichomes (miniature protrusions on the cuticle) that vary among species (McGregor
302 et al. 2007). Thus, work on this one gene has provided us with hypotheses about the
303 function of regulatory and coding regions, some of which are expected to influence
304 many phenotypes, some to have highly targeted phenotypic effects, while some may
305 not have any phenotypic effects. However, note for all this effort, we are far from
306 having any detailed predictions of, for example, what effects that substitution of
307 specific base pairs will have on phenotypes.

308 A complementary tool that is now being exploited to build GP maps is the
309 genome-wide association study (GWAS). In a GWAS, a set of genotypes are
310 sequenced and their phenotypes are measured. When a statistically robust associa-
311 tion between a pair of variants and a phenotype is discovered, we have measured the
312 effect of those variants on a phenotype, one of the crucial parameters of both
313 population and quantitative genetic models (Bastide et al. 2013). Ultimately merging
314 the results of developmental genetic, evo-devo, and GWAS studies coupled with
315 functional verification will allow us to make increasingly detailed GP maps to inform
316 micro-evo-devo studies. An important challenge for GWAS studies is to incorporate
317 pleiotropic effects (Pitchers et al. 2019).

318 Detailed models of development and the phenotypes it produces is a promising
319 approach to building a hypothesis about GP maps (see the chapters ► [“Modeling](#)
320 [Evolution of Developmental Gene Regulatory Networks”](#) and ► [“Computational](#)
321 [Modeling at the Cell and Tissue Level in Evo-Devo”](#)). For example, a detailed model

322 of tooth development can reproduce the major features of tooth morphology and
323 provides hypotheses about which aspects of development will respond under simu-
324 lated selective scenarios (Milocco and Salazar-Ciudad 2020). If such a model were
325 coupled to experimental work in a model organism, this would be a powerful
326 program to build a GP map, and then test its ability to predict micro-evolution.

327 What Can Micro-Evo-Devo Do for Evo-Devo?

328 Given the status of micro-evo-devo as a younger step-child of traditional, compar-
329 ative evo-devo, an important question to ask is what micro-evo-devo can bring to the
330 study of the evolution of development that is distinct from what can be learned using
331 the comparative approach. One entrée to potential advantages of micro studies is to
332 consider the limitations of traditional evo-devo studies.

333 The first such limitation is what we might call the “discreteness bias” of evo-devo
334 studies. The chosen targets of evo-devo studies are overwhelmingly discrete differ-
335 ences among taxa – the transformation of wings to halteres in insects, the evolution
336 of novel color patterns, the loss of limbs in snakes. The ability to investigate such
337 discrete differences is a strength of evo-devo that only becomes a bias when there is a
338 tendency to equate all interesting developmental changes with the causation of
339 discrete differences. Questions about the evolution of important quantitative char-
340 acteristics, such as differences in body size and the accompanying allometric
341 changes in virtually every aspect of form are also interesting. Generalizations
342 about the evolution of form are nevertheless being built on the catalog of causes of
343 discrete differences among taxa (e.g., Carroll 2008).

344 Micro-evo-devo studies generally do not start out to explain discrete phenotypic
345 differences, and thus help balance the scope of evo-devo. For example, Kamberov
346 et al. (2013) studied an amino acid substitution in the human Ectodysplasin A (*Eda*)
347 receptor (*EDAR*). This human polymorphism was targeted for study after population
348 genetic studies revealed that the derived *EDAR* allele has been strongly favored by
349 selection in East Asian and Native American populations over the past 30,000 years.
350 Subsequent studies showed that it was associated with differences in scalp hair
351 thickness and tooth shape in human populations. Kamberov et al. set out to verify
352 that the coding variant was in fact responsible for these phenotypic associations by
353 introducing the amino acid change into the mouse genome. The knock-in mice did,
354 in fact, have 25% more hair follicles whose thickness was enhanced by one cell
355 width, but also revealed 10–25% higher density of eccrine (sweat) glands, increases
356 in branching in the mammary gland by about 25%, and 10%, and lower mammary
357 fat pad size. Subsequent investigations in humans showed that individuals carrying
358 the derived allele also had increased numbers of eccrine glands, as the mouse results
359 predicted. While it remains unclear which of the phenotypic effects of the *EDAR*
360 have been responsible for its spread in this human population, it is a revealing
361 example of positive selection of a highly pleiotropic quantitative genetic variant.

362 Micro-evo-devo studies are well equipped to study the evolution of continuous
363 traits rather than discrete differences. Patterns in continuous traits can be as relevant

364 to larger scale evolutionary patterns as discrete differences. For example, there is
365 long-standing controversy over how much allometry constrains evolutionary pat-
366 terns. Bolstad et al. (2015) performed artificial selection on the slope of a conserved
367 pattern of allometry for wing shape and size in *Drosophila melanogaster*. In just
368 26 generations the slopes evolved to be outside the range of slopes in the entire genus
369 *Drosophila*, suggesting that allometry does not constrain wing shape. However,
370 when artificial selection ceased, the slopes evolved back towards the original slope
371 faster than they diverged under artificial selection. They also selected wing shape to
372 evolve without altering allometric slope, but in those lines, there was little evidence
373 of natural selection for the ancestral state. Consequently, Bolstad et al. interpreted
374 this counter-selection as the result of a pleiotropic burden (see the chapter ▶ “**Con-**
375 **cept of Burden in Evo-Devo**”) on allometry, rather than selection on wing shape per
376 se. This suggests that this allometric relationship is constrained by pleiotropy with
377 unknown aspects of development, not by a lack of genetic variation in slope.

378 It is more challenging to use the micro-evo-devo toolkit to study the opportunity
379 for discrete changes in morphology, as, in many cases, discrete phenotypic variation
380 is not present within populations. Studies of mutational effects can sometimes reveal
381 discrete differences, such as the transformations of cell fate in the vulva of
382 *Caenorhabditis elegans* (Braendle et al. 2010; see the chapter ▶ “**Devo-Evo of**
383 **Cell Types**”). In natural isolates, the pattern of cells is nearly invariant, but after
384 mutations are allowed to accumulate for many generations, this pattern becomes
385 more variable. Mutations in different genotypes generated different rates of trans-
386 formations, and there were hints that the sorts of transformations that occur at the
387 highest rate are more typical of those found in related species. In rare instances,
388 populations contain individuals of two or more discrete morphological types, and
389 these can be used to study the underlying developmental basis.

390 The second potential limitation is “endpoint bias.” Having demonstrated the role
391 of a particular gene in shaping phenotypic differences between taxa, it is natural to
392 equate the evolution of the feature in question to the evolution of that gene. For
393 example, Kopp et al. (2000) demonstrated that the gene *bric a brac* (*bab*) has AU2
394 evolved a novel, sex-specific regulatory role in the last two abdominal segments in
395 some *Drosophila* species. This novel regulation enables male flies to evolve pig-
396 mentation and shape differences from female flies in those segments. Having
397 demonstrated this, Kopp et al. then proposed a parsimonious selective scenario
398 whereby selection on *bab* expression by sexual selection is responsible for the
399 evolution of the novel male-specific features. However, an alternative hypothesis
400 is that the initial evolution of sexual differentiation took place by some other
401 mechanism, and the recruitment of *bab* into its current regulatory role took place
402 later.

403 We do not criticize Kopp et al. for putting forward their parsimonious hypothesis,
404 but there are theoretical and empirical grounds to suspect that the differences in
405 development between species may be different from the mechanisms by which the
406 phenotypes diverged in the first place. On the empirical side, developmental systems
407 drift (DSD) is a well-known and common phenomenon whereby the developmental
408 basis of traits diverges even though the trait has remained roughly constant since

409 divergence from the common ancestor. One potential explanation for this phenom-
410 enon is literally genetic drift – that there are completely equivalent ways to make the
411 same phenotype, and this allows fixation of alleles that transition the system from
412 one developmental solution to another (see the chapter ▶ “Developmental Systems
413 Drift”). Perhaps a more likely explanation for DSD is suggested by Fisher’s geo-
414 metric model (Orr 2005). When the population is far from the optimum, the variants
415 with large effects initially fixed are likely to move some traits farther from their
416 optimal state. Once such an imperfect mutation becomes fixed in the population,
417 mutations fixed at later steps in the adaptive process will rectify the pleiotropic side
418 effects of those early steps. The model predicts that adaptation will tend to zig-zag
419 towards its final state, rather than proceeding by a single fixation event that achieves
420 the optimal state. Thus, the developmental solution to an adaptive challenge may
421 well shift once the population is near the optimum state. For example, a plausible
422 scenario is that the *bab* enhancer now regulating coloration in male abdomens was
423 recruited to correct the side effects of some earlier mechanism of achieving a similar
424 phenotype. Now consider a second adaptive event that primarily involves different
425 traits. The pleiotropic side-effects of that event may perturb development of our focal
426 traits, again engendering evolution to restore that optimum phenotype (Pavlicev and
427 Wagner 2012).

428 How these inferred shifts in the developmental basis of evolved states occur has
429 not yet been addressed by micro-level experiments. Artificial selection and experi-
430 mental evolution studies of developmentally interesting organisms have generated
431 very few allelic fixation events due to their short duration, precluding examination of
432 this issue. Natural experiments such as the repeated invasions of sticklebacks into
433 freshwater habitats have a greater range of time depths and offer the opportunity to
434 evaluate this possibility, although we know of no attempts to do so.

435 Do Micro Events Affect Long-Term Evolution?

436 There are many reasons to suspect that the micro-evolutionary events and processes
437 that we can observe directly are not relevant to evolution above the species level and
438 over long time periods (see the chapter ▶ “Macroevolution”). For example, if the
439 cis-regulatory changes of large effect in fact dominate long-term evolution (Carroll
440 2008), it is possible that the vast majority of the abundant polygenic variation within
441 contemporary populations could have no long-term importance. Some paleontolo-
442 gists have long argued for such a discontinuity between micro-evolutionary pro-
443 cesses and macro-evolutionary patterns on other grounds (Gould 2002). Even
444 quantitative geneticists have generally doubted that the parameters measured within
445 contemporary populations are stable enough to predict evolution over long time
446 periods. Indeed a study that integrates data on evolutionary rates of body size
447 evolution across time scales from generations to hundreds of millions of years
448 suggests a discontinuity reminiscent of punctuated equilibrium (Uyeda et al.
449 2011). Up to a time depth of about one million years, body size of descendant
450 populations or species always remains within a factor of about 1.5 of the ancestral

451 value, but at longer time scales larger changes in body size become increasingly
452 likely. It is not clear what processes can explain this apparent discontinuity, but it
453 certainly suggests that it is very unlikely that one of these rare large changes will be
454 encountered in any particular population at the present. These doubts about rele-
455 vance are likely an important factor in the relative paucity of micro-evo-devo studies.

456 The result, however, is that we have relatively little data we can use to empirically
457 test whether micro- and macro-evolution are in fact related to each other. Recent
458 studies suggest that in some taxa there may be a strong connection between micro
459 processes and macro patterns, despite the many reasons there should not be. Perhaps
460 the most striking example is the recent comparison of the diversification of wing size
461 and shape in the dipteran family Drosophilidae (Houle et al. 2017). This contribution
462 estimated the phenotypic variation produced by mutation in 21 aspects of wing size
463 and shape (Houle and Fierst 2013) in *Drosophila melanogaster* and compared that
464 variation to standing variation in the same species and to variation among over
465 100 Drosophilid species that diversified over at least 30 million years. Both muta-
466 tional and standing variation within *D. melanogaster* were highly correlated with the
467 rate of evolution in the family. Wing shape evolves slowly in this group, so it is
468 possible that this strong relationship is a peculiarity of a suite of slow-evolving traits.
469 However there are hints of similar patterns in traits that show higher rates of
470 evolution as well, such as the blossoms of the plant genus *Dalechampia* (Bolstad
471 et al. 2014). The relevance of micro-evo-devo studies to the broader field of
472 evo-devo should not be lightly dismissed.

473 Cross-References

- 474 ▶ [A Process-Based Approach to the Study of Flower Morphological Variation](#)
- 475 ▶ [Computational Modeling at the Cell and Tissue Level in Evo-Devo](#)
- 476 ▶ [Concept of Burden in Evo-Devo](#)
- 477 ▶ [Developmental Evolutionary Biology \(Devo-Evo\)](#)
- 478 ▶ [Developmental Plasticity and Evolution](#)
- 479 ▶ [Developmental Systems Drift](#)
- 480 ▶ [Eco-Evo-Devo](#)
- 481 ▶ [Epistasis](#)
- 482 ▶ [Evo-Devo's Contributions to the Extended Evolutionary Synthesis](#)
- 483 ▶ [Evolvability](#)
- 484 ▶ [Genotype-Phenotype Map](#)
- 485 ▶ [Macroevolution](#)
- 486 ▶ [Modeling Evolution of Developmental Gene Regulatory Networks](#)
- 487 ▶ [Modularity in Evo-Devo](#)
- 488 ▶ [Phenotyping in Evo-Devo](#)
- 489 ▶ [Variational Approaches to Evolvability: Short- and Long-Term Perspectives](#)

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